



Fundusze Europejskie
dla Rozwoju Społecznego



Rzeczpospolita
Polska

Dofinansowane przez
Unię Europejską



COURSE SYLLABUS		
1.	Course title:	
	Cutting Edge Techniques Advancing Foraminiferal Culture experiments <i>To be agreed individually with the staff and participants, within the period of April to August 2026, Kraków, Poland</i>	
2.	Lecturers (incl. Instructors):	
	<i>Prof. Jarosław Tyszka, PhD, Institute of Geological Sciences, Polish Academy of Sciences</i> <i>Michael Lintner, PhD, Eng., Institute of Geological Sciences, Polish Academy of Sciences</i> <i>Jan Goleń, MSc, Institute of Geological Sciences, Polish Academy of Sciences</i> <i>Joanna Szalkowska, MSc, Eng., Inst. of Geological Sciences, Polish Academy of Sciences</i> <i>Karolina Godos, MSc, Institute of Geological Sciences, Polish Academy of Sciences</i>	
3.	Field, type and level of studies, year of study:	
	<i>Earth sciences, oceanography, paleoecology, marine biology, environmental pollution (bachelor, master, and doctoral students)</i> <i>all years students in area of geology, biology ecology, geography, chemistry or similar</i>	
4.	Course character:	
	<i>Workshop with laboratory work and training sessions</i>	
5.	Teaching method:	
	<i>Lectures, practical exercises, and culture experiments</i>	
6.	Language:	<i>English</i>
7.	Course type and number of hours:	
	Workshop (35h)	
8.	Estimated load of student's independent work:	10 h



**Krótkookresowa wymiana akademicka sposobem na podniesienie
jakości kształcenia w instytucjach szkolnictwa wyższego i nauki**
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9.	Total workload and number of ECTS points:	2 ECTS
10.	Short description and main focus of the course:	
	The primary aim of this 5-day intensive workshop is to significantly enhance the laboratory skills of young scientists engaged in actualistic studies of foraminifera. Participants will gain the tools necessary to address complex biological and geological questions by mastering high-resolution imaging and physiological monitoring. The course bridges the gap between raw observation and analytical depth, focusing on three core research pillars: • Biom mineralization and Morphogenesis: Visualizing how shells are formed and organized at a subcellular level. • Biomonitoring and Ecotoxicology: Assessing the real-time physiological response of foraminifera to environmental stressors or pollutants. • Palaeoenvironmental Reconstructions: Utilizing modern biological observations to improve the calibration of proxies used in deep-time climate studies. Key Thematic Areas & Methodologies • Advanced Fluorescence & Confocal Imaging (FV4000 & AxiObserver Z.1) ◦ Experimental Architecture: Designing protocols that balance signal-to-noise ratios by optimizing laser power, exposure times, and precise detection ranges. ◦ Control imaging: Detection of autofluorescence ◦ Dynamic Staining: Selecting and applying fluorescent dyes to target specific organelles or physiological processes. ◦ Live Imaging Control: Introducing incubation and perfusion chambers to control environmental conditions and active application of reagents. • Physiological & Photosynthetic Assessment ◦ MICROSCOPY-PAM: Utilizing Pulse Amplitude Modulated (PAM) Fluorometry to measure the photochemical efficiency and health of symbiotic algae within Large Benthic Foraminifera (LBFs). ◦ Isotopic Labelling: Introduction to the fundamentals of tracer application to quantify metabolic rates and calcification pathways. • Digital Image Synthesis ◦ Post-Processing & Deconvolution: Transforming raw optical sections into clear, high-contrast data sets by removing "out-of-focus" light blur. ◦ Quantitative Analysis: Moving from qualitative "pretty pictures" to extracting measurable data for statistical scientific reporting.	



11.	References:
	Dubicka Z., Tyszka J., Pałczyńska A., Höhne M., Bijma J., Jense, M., Klerks, N., & Bickmeyer U. (2024). Biocalcification in porcelaneous foraminifera. <i>Elife</i>, 13, RP91568, https://doi.org/10.7554/eLife.91568.3 Frontalini F., Losada M.T., Toyofuku T., Tyszka J., Goleń J., de Nooijer L., Ikuta T. (2019). Foraminiferal ultrastructure: a perspective from fluorescent and fluorogenic probes. <i>Journal of Geophysical Research: Biogeosciences</i>, 124(9), 2823-2850, https://doi.org/10.1029/2019JG005113 Goleń J., Tyszka J., Godos K., & Janse M. (2022). A Model of F-actin Organization in Granuloreticulopodia in Foraminifera: Morphogenetic and Evolutionary Implications from Novel Fluorescent and Polarised Light Observations. <i>Protist</i>, 173(4), 125886, https://doi.org/10.1016/j.protis.2022.125886 Lintner, M., Schagerl, M., Lintner, B., Nagy, M., & Heinz, P. (2022). Photosynthetic performance of symbiont-bearing foraminifera <i>Heterostegina depressa</i> affected by sunscreens. <i>Scientific Reports</i>, 12(1), 2750, https://doi.org/10.1038/s41598-022-06735-1 Lintner, M., Balzano, S., Keul, N., Heinz, P., Manecki, M., Klimek, A., ... & Tyszka, J. (2025). Biosorption of heavy metals by microalgae: Hazardous side effects for marine organisms. <i>Chemosphere</i>, 372, 144080, https://doi.org/10.1016/j.chemosphere.2025.144080 Tyszka J., Bickmeyer U., Raitzsch M., Bijma J., Kaczmarek K., Mewes A., Janse M. (2019). Form and function of F-actin during biomineralization revealed from live experiments on foraminifera. <i>Proceedings of the National Academy of Sciences</i>, 116(10), 4111-4116, https://doi.org/10.1073/pnas.1810394116
12.	Prerequisites:
	Micropaleontology, Biogeochemistry, Marine biology, Paleo/ecology, Environmental Toxicology
13.	Educational outcomes:
	Knowledge: Upon successful completion of this course, students will be able to: • Microscopy Principles: Understanding the operational physics behind fluorescence and confocal microscopy (specifically the FV4000 and AxiObserver Z.1 systems). • Experimental Design: Learning how to select optimal imaging parameters, including laser power, exposure times, and appropriate fluorescent dyes/filter sets. • Physiological Monitoring: Understanding the basics of isotopic labeling and chlorophyll fluorometry (MICROSCOPY-PAM) to assess the metabolic and photosynthetic activity of foraminifera and their symbionts.



	• <i>Environmental Control: Knowledge of using incubation and perfusion chambers to manipulate CO₂, O₂, and temperature during live imaging.</i>
	Practical Skills: <i>This course provides a comprehensive introduction to:</i> • <i>Sample Preparation: Mastering the delicate process of picking, cleaning, and staining living foraminifera.</i> • <i>Advanced Imaging: Hands-on operation of confocal microscopes, including the application of inhibitors and stains during real-time imaging.</i> • <i>Data Processing: Competence in the processing, deconvolution, and quantitative analysis of complex biological images.</i> • <i>Equipment Operation: Direct experience with the MICROSCOPY-PAM system for measuring chlorophyll activity.</i>
	Social Skills: <i>During the workshop, students will:</i> • <i>Collaborate effectively within small research teams</i> • <i>Clearly and constructively present scientific observations in group discussions</i> • <i>Develop critical thinking and problem-solving skills through collaborative interpretation of scientific data</i> • <i>Demonstrate awareness of research ethics, data integrity, and open science principles in team-based research activities</i>
14.	Evaluation of the educational outcomes:
	<i>Students will be evaluated based on their engagement in the workshop, participation in discussions, and performance in the practical exercises</i>
15.	Criteria to complete the course:
	<i>To successfully complete the course, students must: 1) Attend at least 80% of all scheduled classes and lab activities; 2) Actively participate throughout the workshop; 3) Obtain a positive evaluation of practical exercises</i>
16.	Contact with the course leader/lecturer:
	Prof. Jarosław Tyszka, ndtyszka@cyfronet.pl Dr Michael Lintner, ndlintne@cyfronet.pl Jan Goleń, MSc, ndgolen@cyfronet.pl